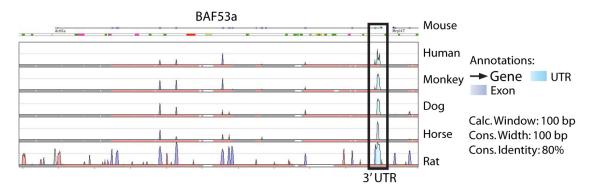
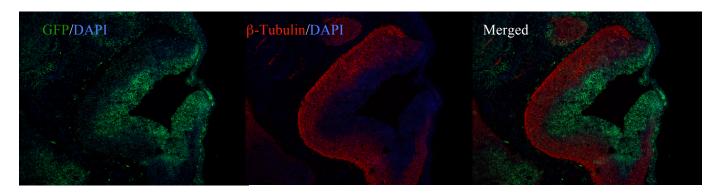
doi: 10.1038/nature08139

Supplementary Figure 1



Cross-species alignment of BAF53a genomic regions of mammals by VISTA1.

The peaks represent conserved sequences in a minimal 80% identity in a scanning window of 100 base pairs. The peaks are present at the coding regions while the most conserved region resides in the 3' UTR (shown by the light blue peaks in the rectangular enclosure). The genomic region shown above is approximately 18 kb.

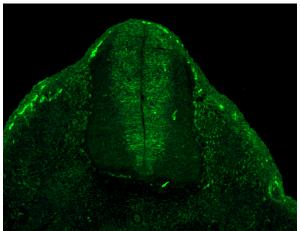


BAF53a BAC d2nucEGFP-BAF53a WT UTR

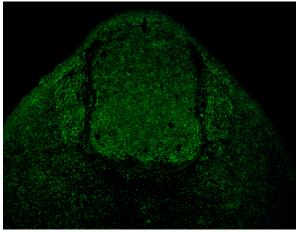
Sagittal sections of the E11.5 brain of BAF53a BAC reporter near the 4^{th} ventricle. Downregulation of the BAF53a reporter expression is consistent in β -Tubulin-positive neurons in the brain, demonstrating the pan-neuronal specificity of BAF53a repression.



Cross-species alignment of 3' UTRs of BAF53a. The conserved regions corresponding to the predicted binding sites of miR-9, miR-9* and miR-124 are shown in highlights in yellow, green and blue, respectively. The predicted configurations of the microRNAs binding to target sites in the 3' UTR of BAF53a are shown in Fig. 2a.

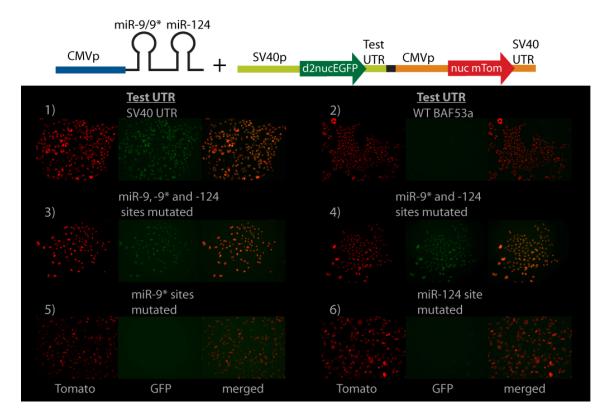






BAF53a BAC d2nucEGFP-BAF53a **MUT** UTR

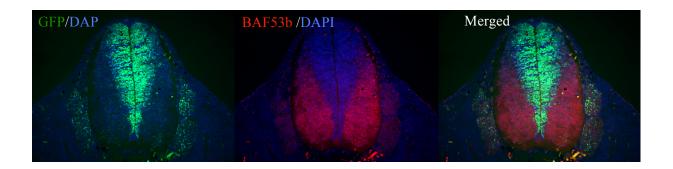
Repression of BAF53a BAC reporter is specific for differentiated neurons. Top panels show wide views of GFP signals from the cross sections of transgenic embryos for the wild type BAF53a BAC reporter and the mutant BAF53a BAC reporter (containing mutations in BAF53a 3'UTR of d2nucEGFP). The de-repression of the reporter expression seen with the mutant reporter (right panel) appears to be specific for differentiated neurons in the neural tube and the neural crest-derived neurons.



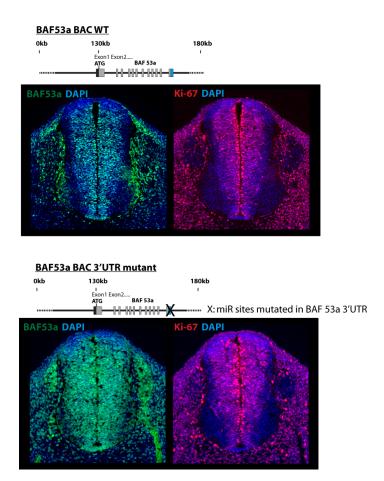
Schematic diagram of microRNA expression and sensor constructs. We synthesized a cluster of miR-9* and miR-124 precursors driven by CMV promoter (top diagram). The production of mature miR-9* and miR-124 was confirmed by quantitative RT-PCR (data not shown). The sensor (d2nucEGFP) and marker (nuclear Tomato) reporters were driven by SV40 and CMV promoters, respectively. The photographs show representative fields of stable CHO lines that contain the microRNA overexpression and sensor constructs (quantified in Fig.2c). Note that stable CHO lines showed significantly reduced EGFP (sensor) expression with 3' UTR of BAF53a. This EGFP downregulation was abolished with mutations in miR-9* and miR-124 binding sites.

Test UTRs: 1) SV40 3' UTR, 2) wild type BAF53a UTR, 3) BAF53a 3' UTR with mutations in miR-9, -9* and -124 sites, 4) mutations in miR-9* and -124 sites, 5) mutation in miR-9* site only, and 6) mutation in miR-124 site only.





Overexpression of miR-9*/-124 in neural progenitors and reduction of BAF53a does not activate BAF53b expression in E11.5 transgenic embryos. The top drawing represents a schematic diagram of the miR-9*/-124 expression construct. GFP-positive progenitors show decreased population of BAF53a and Ki-67-positive cells, indicative of proliferative defects (Fig. 4b). Although the progenitors exit from cell cycle, there appears to be no overlapping expression between GFP and BAF53b as shown in the photos.



Ki-67 expression in wild type and mutant BAF53a BAC-transgenic embryos.

The top panel shows E11.5 embryos transgenic with wild-type BAF53a BAC which showed normal downregulation of BAF53a and a lack of Ki-67 expression (a marker for proliferation) in the neuronal zone. Mutating the 3' UTR of BAF53a lead to extended expression of BAF53a in E11.5 transgenic embryos. Extended expression of BAF53a was associated with disorganized mitosis and extended Ki-67 expression. This phenotype may be related to the possible expression of miR-9* (Fig. 2d) and miR-124² along the ventricular surface of the neural tube, which has been proposed to a site of neurogenic asymmetric division of progenitors to produce neurons ³⁻⁵.

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